Synthesis of the Antibiotic (*R***)-Reutericyclin** *via Dieckmann* **Condensation**

by **Roswitha Böhme**a), **Günther Jung***b), and **Eberhard Breitmaier**a)

a) Kekulé Institute of Organic Chemistry and Biochemistry, University of Bonn, Gerhard-Domagk-Strasse 1, D-53121 Bonn (e-mail: info@ebmaier.de) b) *EMC microcollections GmbH*, Sindelfinger Strasse 3, D-72072 Tübingen

(e-mail: emc@microcollections.de)

(*R*)-Reutericyclin ((*R*)-**1**), a bactericidal, amphiphilic natural product with a trisubstituted tetramic acid moiety, was prepared in four steps from D-leucine in an overall yield of 24%. The chiral heterocyclic portion of **1** was synthesized by *Dieckmann* cyclization of ethyl *N*-(acetoacetyl)leucinate (**7**), and the resulting pyrrole derivative 8 was *N*-acylated with (E) -dec-2-enoyl chloride in the presence of BuLi at -70° (*Scheme 2*). This new procedure is straightforward and allows the synthesis of both antipodes of reutericyclin in an enantiomeric excess (ee) of *ca*. 80%.

Introduction. – Reutericyclin $(=(2R)$ -4-acetyl-1,2-dihydro-5-hydroxy-2- $(2$ -methylpropyl)-1-[(2*E*)-1-oxodec-2-enyl]-3*H*-pyrrol-3-one; (*R*)-**1**)1) is a tetramic acid that acts against a broad spectrum of *Gram*-positive bacteria. The amphiphilic metabolite was the first low-molecular-weight antibiotic isolated from a lactic acid bacterium, *i.e*., *Lactobacillus reuteri* LTH2584 [1][2]. Reutericyclin is produced during food industrial sourdough fermentation by *Lactobacillus reuteri*, and contributes to the resistance of its producer strain [3]. The mode of action is based on its proton-ionophoric properties, selectively dissipating the transmembrane proton potential (ΔpH) [4] [5].

There are two general pathways to synthesize reutericyclin (**1**). In the first published synthesis [6], leucine *tert*-butyl ester is *N*-acylated with (*E*)-dec-2-enoyl chloride to the amide **2** (*Scheme 1*). After ester hydrolysis, the resulting *N*-acylamino acid is reacted with *Meldrum*'s acid to the intermediate condensation product **3**, which thermally cyclizes to the tetramic acid **4**. The Ac group is introduced during the last of the seven steps (starting from leucine) to yield racemic **1**. Racemization is due to the last

¹⁾ Systematic name of the *enolic* form drawn (tautomerism; see *Figure* in the *Exper. Part*).

^{© 2005} Verlag Helvetica Chimica Acta AG, Zürich

step in which TiCl4, a strong *Lewis* acid, is used as a catalyst. The separation of the enantiomers of racemic reutericyclin is expected to be difficult for large-scale preparations. Furthermore, the reaction conditions for the thermal cyclization with *Meldrum*'s acid have to be strictly followed to obtain reproducible results. For these reasons, we developed a second synthetic pathway aiming at the synthesis of the two antipodes of **1**.

Results and Discussion. – To avoid the vigorous conditions typically required for *C*acylation, we designed a synthetic route that includes *N*-acylation with (*E*)-dec-2-enoyl chloride in the *last* step. As outlined in *Scheme 2*, the Ac group was introduced first by coupling D-leucine ethyl ester (**5**) with *N*-hydroxysuccinimidyl acetoacetate (**6**) [7]. The resulting leucinate **7** was cyclized to the 1,2-dihydro-3*H*-pyrrol-3-one **8** *via Dieckmann* condensation in the presence of EtONa/EtOH, following a modified procedure to cyclize pyrrole derivatives [8]. Alternative attempts to introduce the acetoacetyl group and to carry out the *Dieckmann* cyclization failed.

The last, crucial step involved *N*-acylation of tautomeric **8** (*cf*. *Figure*) with (*E*)*-*dec-2-enoyl chloride. Since 8 is a *cyclic* amide, several bases such as DMAP, Et₃N/*N*,*N*dimethylaniline (*via* an Me₃SiN group), pyridine (*Schotten–Baumann* conditions), and $LiNH₂$ were tested as acylation catalysts. However, no coupling product with the acid chloride was obtained. Similarly, when (*E*)-dec-2-enoic acid was activated with 1-hydroxy-1*H*-benzotriazole (HOBt) or transformed into a mixed anhydride with isobutyl chlorocarbonate (using $E(t_i-Pr)$). Or DBU as base), no reaction to 1 was observed. Attempted condensations by fusion of the neat compounds or in the presence of 18-crown-6 in dimethoxyethane (DME) were also unsuccessful. Finally, BuLi (2.2 equiv.)²) was found to remove the amide proton of **8** at -70° under salt formation, and, after 20 min of vigorous stirring, the resulting strong nucleophile was reacted with dec-2-enoyl chloride. After aqueous workup and column chromatographic purification, (R) -1 was obtained as an amorphous powder.

Further purification was achieved by recrystallization. Several solvents were tested to this end, especially EtOH, $Me₂CO$, AcOEt, Et₂O, MeOH, and petroleum ether.

²⁾ Only few cases are known in which unbranched BuLi was used in combination with NH compounds [9].

Scheme 2. *Synthesis of (*R*)-***1** *from Ethyl* D-*Valinate*

a) CH₂Cl₂, r.t., 3 h. *b*) EtONa/EtOH, Δ , 3 h. *c*) 1. THF, BuLi (2.2. equiv.), -70° ; 2. (*E*)-dec-2-enoyl chloride; $3. H₃O⁺$.

Since reutericyclin (**1**) possesses pronounced amphiphilic properties due to its lipophilic alkyl chain and the hydrophilic tetramic acid part, it is soluble in many solvents. In most of the above solvents, it formed a gel upon cooling, rather than crystals; and slow evaporation of the solvent rendered an oily residue. Nevertheless, we finally achieved to obtain a solid material from a cyclohexane/acetone mixture upon slow evaporation.

We observed that the NMR spectra of reutericyclin (**1**) seem to depend on the workup procedure and NMR solvent, which may give rise to different tautomeric forms and H-bonding patterns. Due to staple effects, the ¹H NMR spectra of some reutericyclin preparations showed only poorly resolved signals, with almost no fine structures. However, it was possible to record acceptable ¹H- and ¹³C-NMR spectra and to perform a HMQC experiment with a dilute (D_6) DMSO solution. The complete NMR data of **1** have been previously published [1] [6].

After total hydrolysis and derivatization of the samples of synthetic **1**, GC/MS analysis on a chiral capillary column showed an enantiomeric purity of *ca*. 90% (80% ee) of the corresponding L- or D-leucine, depending on the configuration of the original substrate **5**.

Experimental Part

General. M.p.: *Büchi SMP-20* apparatus; uncorrected. UV Spectra: *Beckmann DU660* spectrometer; *l*max (*e*) in nm. Optical rotation: *Perkin-Elmer 341* polarimeter. IR Spectra: *Perkin-Elmer 1600* spectrometer; in cm⁻¹. NMR Spectra: *Bruker DPX-400* and *DRX-500* spectrometers; at 400 or 500 (¹H), or at 100 or 125 MHz (13C), resp. HR-EI-MS: *Kratos MS-50* spectrometer (*A.E.I*., Manchester).

Figure. *¹ H- and 13C-NMR* (italics) *chemical shifts and multiplicities for compound* **8** *present in two major tautomeric forms*. The observed ratio of A/B was *ca.* 4:1 in CDCl₃ solution at 20°. The two Me groups of the i-Bu side chain were tentatively assigned.

*(*E*)-Dec-2-enoic Acid*. This compound was prepared through *Knoevenagel* condensation of octanal and malonic acid [6] [10]. Variations of temp., base, and reaction time were tested, but always a certain amount of β -hydroxydecanoic acid was detected, which could not be separated completely from the desired product. A most-useful old variant [11] proved to be stirring at r.t. for 36 h. Yield: 70% (for 0.2 mol).

*(*E*)-Dec-2-enoyl Chloride.* This compound was prepared by heating the acid with SOCl2 followed by distillation [6]. Yield 76% (for 0.12 mol).

Ethyl ^D*-Leucinate Hydrochloride* (**5**). D-Leucine was heated under stirring in EtOH soln. saturated with HCl gas [12]. Yield: 87% (for 38 mmol). $[a]_D^{20} = -18$ ($c = 5$, EtOH; $[a]_D^{20}$ of commercially available L-enantiomer: $+18$).

Ethyl N-(*Acetoacetyl*)-*D-leucinate* (**7**) [7]. To a soln. of **5** (4.892 g, 25 mmol) in CH₂Cl₂ (40 ml) was added 2 equiv. of Et(i-Pr)2N (6.463 g, 8.56 ml, 50 mmol). Under Ar gas, a previously prepared suspension of commercial N -(hydroxysuccinimidyl)acetoacetate $(6; 4.979 \text{ g}, 25 \text{ mmol})$ in CH₂Cl₂ (40 ml) was added through a dropping funnel within 15 min, and the resulting mixture was stirred at r.t. for 3 h. The soln. was washed with 5% aq. HCl (5×40 ml), and the aq. layers were re-extracted with CH₂Cl₂ (3×50 ml). The combined org. layers were washed with sat. NaHCO₃ soln. (3×) and H₂O (1×). After drying (Na₂SO₄) and evaporation of the solvent, a pale yellow oil was obtained, which was used without further purification. Yield of **7**: 5.71 g (94%). *M*^r 243.30 g/mol. ¹ H-NMR (CDCl3): 0.93, 0.94 (2*d*, 2 Me of i-Bu); 1.27 (*t*, *Me*CH2O); 1.55 – 1.66 (*m*, CH2 of i-Bu)); 1.68 (*m*, Me2C*H*); 2.27 (*s*, Ac); 3.43 (*s*, AcC*H*2); 4.18 (*q*, OCH2); 4.59 (*td*, NCH); 7.17 (*d*, NH). 13C-NMR $(CDCI₃)$: 14.2 (*MeCH₂O*); 22.0, 22.9 (2 Me of i-Bu); 25.0 (Me₂CH); 31.0 (Me of Ac); 41.4 (CH₂ of i-Bu)); 49.7 (AcCH₂); 51.1 (NCH); 61.4 (OCH₂); 165.4 (COO); 172.7 (NCO); 204.1 (C=O of Ac).

*(2*R*)-4-Acetyl-1,2-dihydro-5-hydroxy-2-(2-methylpropyl)-3*H*-pyrrol-3-one* (**8**) 1). To a soln. of EtONa, prepared by dissolving elemental Na (0.58 g, 1.07 equiv.) in anh. EtOH (18 ml) under Ar, **7** (5.71 g, 23.5 mmol) in EtOH (30 ml) was added through a dropping funnel within 25 min at r.t., and the resulting mixture was refluxed for 3 h. After cooling down, the mixture was neutralized with 10% AcOH in EtOH (4.57 ml), and the solvent was evaporated. CH₂Cl₂ (25 ml) was added to the pale-yellow, gel-like residue, which was washed with H₂O (4 ×) to remove AcONa. The aq. layers were extracted with CH₂Cl₂ (3 ×), the combined org. layers were dried (Na₂SO₄), and the solvent was removed on a rotary evaporator. The remaining solid was crystallized from EtOH in several fractions to afford colorless 8. Yield: 3.152 g (68%). M_1 197.23 g/mol. M.p. 133-134°. UV (EtOH): 215 (60130), 275 (146540). [*a*]_{D}² = +117[°] (*c*=0.1, EtOH)³). IR (KBr): 3226 (NH), 2960 (CH), 1669 (C=O), 1449 (NH), 1367, 1319, 1277, 1229. ¹H- and ¹³C-NMR: see the *Figure*. EI-MS: 197.1 (1.2, *M*⁺), 182.1 (5.5, [*M* – CH₃]⁺), 154.1 (25, [*M* – Ac]⁺), 141.1 (100, [*M* – C₄H₈]⁺), 123.1 (19, [*M* – C₄H₈–H₂O]⁺), 86.1 (30), 70.1 (20).

*(*R*)-Reutericyclin* (= *(2*R*)-4-Acetyl-1,2-dihydro-5-hydroxy-2-(2-methylpropyl)-1-[(2*E*)-1-oxodec-2-enyl]- 3*H*-pyrrol-3-one*; (*R*)-**1**). A 250-ml three-neck round-bottom flask equipped with a strong mechanical stirrer was dried and evacuated several times, flooded with Ar gas, and then filled with a soln. of **8** (2.958 g, 15.00

³⁾ L-Enantiomer: $[\alpha]_D^{20} = -128^\circ$ (EtOH).

mmol) in anh. THF (100 ml). At -70° , 2.2 equiv. of BuLi (1.6M soln. in hexane; 21 ml, 33 mmol) was added through a dropping funnel within 20 min under vigorous stirring. The soln. became turbid as a salt precipitated, first yellowish, then light orange, and stirring was continued for 20 min. Then, 0.95 equiv. of *dec-2-enoyl chloride* (2.83 g, 2.79 ml) in THF (20 ml) was added dropwise within 15 min at -70° . The precipitate gradually disappeared, and the soln. was stirred for another 30 min at this temp. After removing the cooling bath, the mixture was stirred for another 2 h, during which time the soln. reached r.t. Ice water (150 ml) with conc. H_2SO_4 (3.234 g, 33 mmol) was slowly added, and the aq. layer was separated and extracted with CH₂Cl₂ ($3\times$ 50 ml). The org. layers containing THF were diluted with CH₂Cl₂ (70 ml), and then washed neutral with H₂O (4×70 ml). After drying (Na_2SO_4) and evaporation of the solvents, an oily, slightly yellow residue was obtained (4.94 g), which was purified by column chromatography (CC) on silica gel *60 PF254* (*Merck 7747*) with cyclohexane/Me2CO 3 : 1 (column: 12 cm × 6 cm (i.d.)). During chromatography, the yellow substance developed a red ring (probably due to decomposition of a byproduct). Of the three major fractions, the first one was resubjected to CC (conditions as above; column: $15 \text{ cm} \times 3.5 \text{ cm}$ (i.d.)); the other two fractions contained the pure product (TLC, HPLC). Yield of 1: 1.985 g (38%). M_r 349.46 g/mol. M.p. 161-164° (dec.). UV (EtOH): 228 (17870), 266 (23875), 278 (25165). Optical rotation (EtOH, $c=0.29$): $\left[\alpha\right]_{436}^{20} = +69$, $\left[\alpha\right]_{546}^{20} = +19$, $\left[\alpha\right]_{589}^{20} = +15$; $\left[\alpha\right]_{589}^{20}$ $([a]_D)$ =+13°. IR (KBr): 2927 (Me, CH₂), 2856 (CH), 1718 (ketone C=O), 1622 (amide C=O), 1470, 1355, 1319, 1242, 1155, 980 (HC=CH), 761. ¹ H- and 13C-NMR: see [1] [6]. EI-MS: 349.2 (7.5, *M*⁺), 293.1 (42, $[M - C_4H_8]^+$, 250.1 (18, $[M - Ac]^+$), 198.0 (20, $[M + 2 - C_{10}H_{17}O]^+$), 183.1 (25), 153.2 (71, $C_{10}H_{17}O^+$), 141.1 (100) , 69.1 (33) 55.1 (29) . HR-EI-MS: 349.2244 $(M^+, C_{20}H_{31}NO_4^+$; calc. 349.2235).

*(*S*)-Reutericyclin*. This enantiomer was prepared as described for (*R*)-**1**, but from (less-expensive) ethyl Lleucinate. The enantiomeric purity of **1** was determined as follows: a sample of synthetic (*S*)-(**1**) from a batch of *ca*. 2 g of product was hydrolyzed with 6N DCl/D₂O at 110° for 24 h. The dried hydrolysate was esterified with DCl/EtOD at 110° for 30 min, and the resulting ester was *N*-protected with trifluoroacetic anhydride to afford *N*-trifluoroacetyl-D-leucine ethyl ester. GC/MS Analysis of this derivative on a capillary coated with *Chirasil-Val* [13] showed two fully separated peaks (Δt_R 2 min) corresponding to a D/L mixture of 9.7:90.3 (80.6% ee).

Financial support from *EMC microcollections GmbH*, Tübingen, Germany, is gratefully acknowledged. We thank *Graeme Nicholson* for chiral analysis of reutericyclin.

REFERENCES

- [1] A. Höltzel, M. G. Gänzle, G. J. Nicholson, W. P. Hammes, G. Jung, *Angew. Chem., Int. Ed.* **2000**, *39*, 2766.
- [2] M. G. Gänzle, A. Höltzel, J. Walter, G. Jung, W. P. Hammes, *Appl. Environ. Microbiol.* **2000**, *66*, 4325.
- [3] M. G. Gänzle, R. F. Vogel, *Int. J. Food Microbiol*. **2002**, *80*, 31.
- [4] M. G. Gänzle, R. F. Vogel, *Appl. Environ. Microbiol*. **2003**, *69*, 1305.
- [5] M. G. Gänzle, *Appl. Microbiol. Biotechnol*. **2004**, *64*, 326.
- [6] U. Marquardt, D. Schmid, G. Jung, *Synlett* **2000**, *8*, 1131.
- [7] A. W. Trautwein, R. D. Süßmuth, G. Jung, *Biorg. Med. Chem. Lett.* **1998**, *8*, 2381.
- [8] A. Treibs, A. Ohorodnik, *Liebigs Ann. Chem.* **1958**, *611*, 139; H. Bauer, *Liebigs Ann. Chem.* **1970**, *736*, 1; G. Pfeiffer, H. Bauer, *Liebigs Ann. Chem.* **1976**, 383.
- [9] C. Casnati, M. R. Langella, A. Ricca, A. Umani-Ronchi, *Tetrahedron Lett.* **1964**, *24*, 1597.
- [10] F. Bohlmann, H. Bornowski, *Chem. Ber.* **1961**, *94*, 3189.
- [11] V. J. Harding, C. Weizmann, *J. Chem. Soc.* **1910**, *97*, 299.
- [12] R. D. Gillard, R. Wootton, *J. Chem. Soc. B* **1970**, 364.
- [13] H. Frank, G. J. Nicholson, E. Bayer, *J. Chromatogr.* **1978**, *167*, 187.

Received July 11, 2005